

Internal use only Pathology # Mosaic task #

ACS Diagnostic & Research Histology Laboratory Services Submission Form

Centre for Comparative Medicine | 4145 Wesbrook Mall, Vancouver, BC V6T 1W5

Email completed form to acs.lab@ubc.ca

SUBMITTER INFORMATION					
Submitter name:	Facility / Company:				
Submitter phone:	Investigator:				
Submitter email:	Workday Program/Grant/Project/Gift:				
Submission date DD-MM-YYYY:	Bill to:				

	DIAGN	OSTIC SERVICES Jana H Complete sections 1-5	odasova				RESE	ARCH HISTOLO Complete sect	GY Ingrid Barta ions 6-12	
1			ANI	MAL(S	S) / SAMPLE(S)	SUBMIT	TED			
Samp	le type description:			Anim	al(s) submitted:		□ Dead	Fixative (histo	pathology):	
				Anim	iai(s) submitted.	🗆 Live		□ 10% Neutra	l buffered formalin	
				Eutha	anized:	□ Yes	🗆 No	□ Other (spec	cify):	
# of s	samples:			Eutha	anasia method:			Method of urin	e collection:	
								□ Free catch	🗆 Tab	e top / Other
								□ Manual exp	ression 🛛 Urin	ary catheter
								Cystocentes	is	
Samp	ole submitted: 🛛 🗆	Fresh 🗆 Fixed [□ Frozen	Date	of euthanasia:			Testing schedu	le (sentinel only):	Protocol #:
Date	of sample collection:			Anim	al(s) found dead:	□ Yes	□ No			
2			Α	ΝΙΜΑ	L SAMPLE INFO	RMATIO	N			
#	Animal ID* #	Sample ID	Room	#	Rack #		Species / S	train	DOB (DD-MM-Y	r) Sex (F / M)
1										
2										
4										
5										
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7										
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9										
10										

* Mosaic database users must provide Mosaic Animal ID.

3 TESTING LABORATORY							
□ ACS Diagnostic Laboratory □ Anir	nal Health Centre		IDEXX BioAnalytics	□ Other			
4 TESTING REQUESTED							
Sentinel comprehensive testing	Bacteriology		Parasitology	Pathology			
 □ Live comprehensive examination □ Comprehensive examination □ Aerobic culture with antimicrobial suscept □ Anaerobic culture with antimicrobial suscept □ Urinalysis □ Urinalysis with cultur antimicrobial suscept □ Other (specify): 		n ID and otibility vith ID ure, ID and	 Ectoparasites Endoparasites Fecal flotation Fecal egg count 	□ Necropsy (non-sentinel animal) □ Histopathology			
IDEXX BioAnalytics / Charles River Lat	oratories:	External veterinary diagnostic laboratory (IDEXX, AHC, CRL, AHL, other):					
 □ Mouse serology □ Rat serology □ Rat PCR □ Other (specify) 		 □ Hematology □ Clinical chemistry □ Coagulation 	□ Serology □ Virology □ Bacteriology	 Molecular diagnostics Toxicology Other (specify) 			
Testing panel(s) / Individual test(s):							
Other tests / Special requests:							



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5	ANIMAL HISTORY / CLINICAL INFORMATION
Were a	ny abnormalities, clinical signs or unexpected findings (including animal found dead) observed? If yes, describe them in detail and include duration and dates:
vvas th	is an experimental animal or did submitted tissue originate from experimental animal? If yes, specify type and duration of experiment and include dates:
Was ar	y procedure / surgery / manipulation performed? If yes, specify and include site and dates:
Was ar	ny substance or treatment administered? If yes, specify and include route, regime, and dates:
Was ar	y mortality or morbidity noted in the colony or in the room? Was any deleterious phenotype noticed in the colony? If yes, specify and include dates:
ls there	anything in particular you are concerned about or looking for?
Include	any additional information, details or requests:
menuue	

6	6 FIXED TISSUE SPECIMEN INFORMATION						
Species:							
# of samples:	_Wet tissues		Cassettes		Paraffin blocks	Slides for staining	
Current storage medium	:	□ 10% NBF	🗆 70% Ethanol	□ Other (specify))	Time in current storage medium:	
Fixative used, if different	from above:	□ 10% NBF	□ 4% PFA	\Box Other (specify))	Time in fixative:	
Additional information (Additional information (e.g. light sensitivity, known pathologies, etc.):						
7			TISSUE P	ROCESSING REQU	JESTED		
Check all applicable boxe							
□ Processing & embeddi				ly #		bedding only #	
□ Pre-trimming tissue (a Orientation of tissue(s) a				embedding #		d decalcification #	
8					MICAL STAINING		
	thickness (stan					s require a different combination of slides, use	
Set 2 & Set 3 fields as ne	cessary.						
	Set 1: Sample	#t	o	Set 2: Sample #	to	Set 3: Sample # to	
Section thickness (µm)							
No stain							
H&E							
Masson's Trichrome							
Picrosirius Red							
Special sectioning instru	ctions (e.g. seria	I sections, leve	ls):				
9 WHOLE SLIDE IMAGING (Olympus VS120 Slide Scanner)							
9 WHOLE SLIDE IMAGING (Olympus VS120 Slide Scanner) Contact the lab for more details and if higher magnifications are required.							
Number of slides for: 10X mag 20X mag 40X mag Image delivery: USB (client to provide) UVD							
10 PATHOLOGY SERVICES							
Number of slides to revie			□ Pathology rep			topathological scoring required*	

* Contact the lab for more details.

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11 SPECIMEN IDENTIFICATION							
Please keep sample IDs succinct as cassette and slide labelling software limit each field to 6 characters. For samples with more than one tissue type, enter each tissue on a separate line. Attach additional lines if necessary. It is not required to complete all columns. If preferred, the same information can be submitted in an attached							
spreadsheet. Sample #	Experiment / Sample ID	Animal ID*	Genotype	Tissue type			
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	ic Animal ID where applicable.						

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